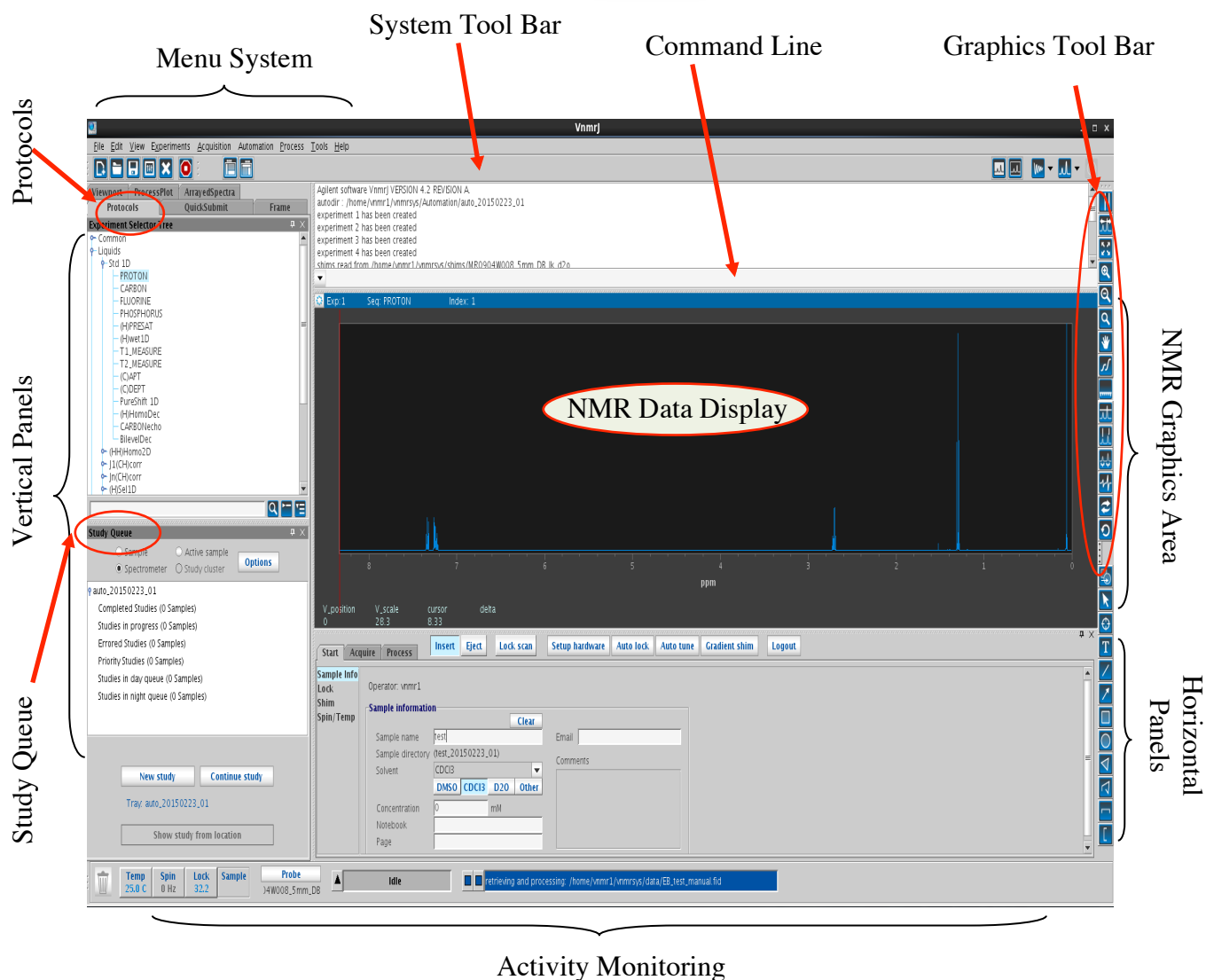




Varian Solution NMR Automation Procedure

(VNMRJ 4.2 under Red Hat Enterprise 5.1)

Jerry Hu, x7914, jghu@mrl.ucsb.edu
 Shamon Walker, x3248, shamonwalker@mrl.ucsb.edu
 Materials Research Laboratory, UCSB
 Version 1.2, Last Modified Wednesday, Sept. 25th, 2017

*****Safety Issues*****

- ⚠ If you, or people working with you, have magnetic metal implants, please consult your doctor for possible effect of magnetic field;
- ⚠ For those who have pacemakers, please stay away from NMR magnets;
- ⚠ Remove from your pocket anything ferromagnetic or vulnerable to magnetic field:
 - Your wallet, bank cards, credit cards, and any cards with magnetic stripes;
 - Electronics: cell phone, mp3, ipod, etc.;
 - Mechanic watches;
 - Keys and other magnetic items.

Table of Contents

<i>I. Facilities Billing System (FBS)</i>	<i>4</i>
<i>II. Sample preparation</i>	<i>5</i>
<i>III. Sign onto Logsheet</i>	<i>5</i>
<i>IV. Start VNMRJ Software</i>	<i>5</i>
<i>V. Preparation: Load Samples into Auto-Sampler</i>	<i>6</i>
<i>V. 1D ¹H and ¹³C NMR Setup and Data Acquisition</i>	<i>8</i>
<i>VI. Finishing up</i>	<i>14</i>
<i>VII. Data Processing Workstation</i>	<i>15</i>
<i>VIII. NMR Data Processing</i>	<i>15</i>
<i>IX. Appendices</i>	<i>21</i>
1. Requirements for Access to the MRL NMR at CNSI	21
<i>X. NMR Basic Principles</i>	<i>22</i>

I. Facilities Billing System (FBS)



To Schedule time and to use the instrument computer a FBS account is required. After training you will receive an invite for FBS or if you already have a FBS account the 400MHz DNP calendar will be added to it.



The instrument time is billed through FBS. Your FBS time = your recharged time. Recharge is calculated at an hourly rate. For current recharge rates go to the MRL website, http://www.mrl.ucsb.edu/sites/default/files/mrl_docs/rechargerate.pdf

- ❖ To begin an instrument session, you must log onto FBS first and either click on **Start Timer** if a reservation has already been made or select **Walk-Up**.

To do so use the FBS designated computer in the lab or any internet connected device, and navigate to <http://ucsb.fbs.io>

- ❖ Now you can log onto your account on the instrument computer. Remember to log off your computer account when finished.
 - ❖ Once the session is finished you must log into your FBS account and click **Stop Timer**. This will stop your FBS time. If you do not do this you could incur extra charges.
 - ❖ The paper log sheet by the instrument is used as back up for the FBS system. Remember to mark you time, recharge number, and any notes or problems you would like to convey, feel free to use as many lines on the page as needed to be clear.
-
- ⚠ No shows will be charged **75%** of the scheduled time. If you cannot use your reserved time cancel it.
 - ⚠ FBS records the billable time as the longer time between the scheduled time and the time used. So if your scheduled time is longer than the actual time on the instrument then the scheduled time will be charged.

II. Sample preparation

1. NMR tubes of **5mm** in OD and **7"** long are used and available from:

- i. Chemistry Department Stockroom, UCSB (Phone: x2107)
- ii. Aldrich
- iii. Wilmad
- iv. Newera



Please get tubes for 600MHz or higher (500MHz grade tubes will work).

2. Samples are dissolved in **deuterated** solvents (~0.1 mM and 50 mM in concentration for ^1H and ^{13}C , respectively, and at least 0.6 mL in volume or 56 mm in height) for three purposes:
 - i. Deuteration removes solvent ^1H signals which would otherwise dominate the ^1H spectrum.
 - ii. Deuterons provides a lock signal.



Lock is a deuterium NMR process that the spectrometer uses to prevent the magnetic field from changing during the course of NMR experiments, thus locking the spectrometer.

- iii. Deuterons provide chemical shift reference for NMR spectra, rendering addition of reference standards such as TMS unnecessary.



**Label your samples with your name and your advisor's name.
This helps us take care of unknown samples.**

III. Sign onto Logsheet

Enter

1. your name
2. your advisor's name and department
3. your recharge account number (in the format: 8-4xxxxx-xxxxx-3)
4. your start time
5. **(Do this at the end of experiment:** your stop time and duration of experiment)
6. **(Do this at the end of experiment:** Status of instrument and report problems if any)

IV. Start VNMRJ Software

1. Make sure that the spectrometer is idle by looking at the computer. If yes, proceed to Step 2 below (if no, either wait, talk to the user, or do something else)
2. Login into your NMR account:
Type your username, hit the Enter key

Type your password, hit the Enter key again



3. Double click on the VNMRJ icon on the desktop, the last VNMRJ layout from your previous login session will appear.

V. Preparation: Load Samples into Auto-Sampler

1. Put your sample in a green spinner , measure depth with the golden depth gauge  or the rack for spinners, and clean the bottom half of the sample tube with a napkin while holding the top half of the tube;



Spinner

Sample should cover the opening,
min. 0.7ml

2. (**Important information**) Slot 12, aligned with the yellow triangle label at the bottom front of the sampler, is reserved for the standard idle sample (pure CDCl_3). **Don't load any sample into this slot.**



The standard idle sample will be loaded into the magnet **automatically** at the end of an automation run.

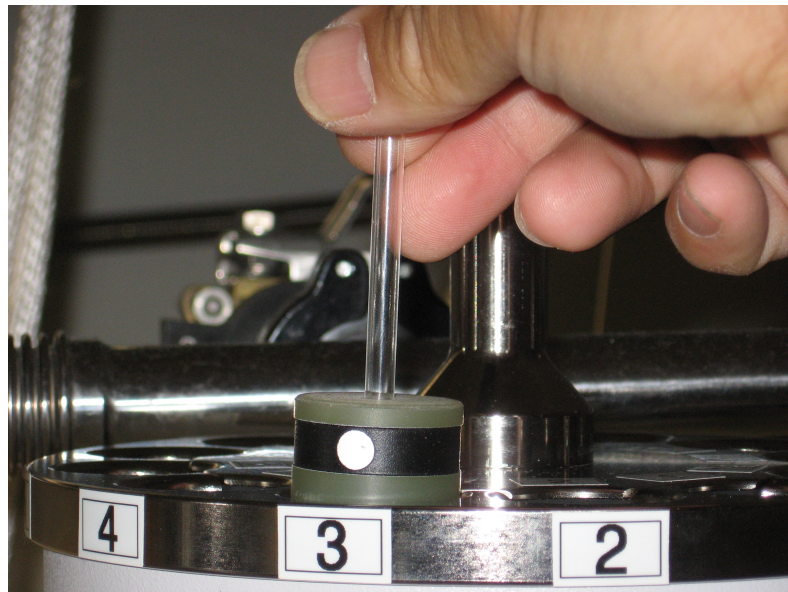


Front View



Top View

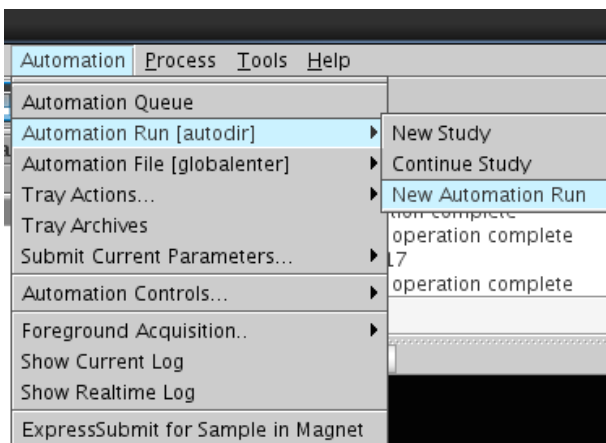
3. Go to the magnet and load your samples onto the sampler in the clockwise order: the 1st sample goes to slot 1, the 2nd to 2, the 3rd to 3, and so on.



 Be very careful to align the sample tube with the slot holes when removing or loading samples.

V. 1D ^1H and ^{13}C NMR Setup and Data Acquisition

1. Choose **New automation run** from the **Automation** menu to create a new automation session.

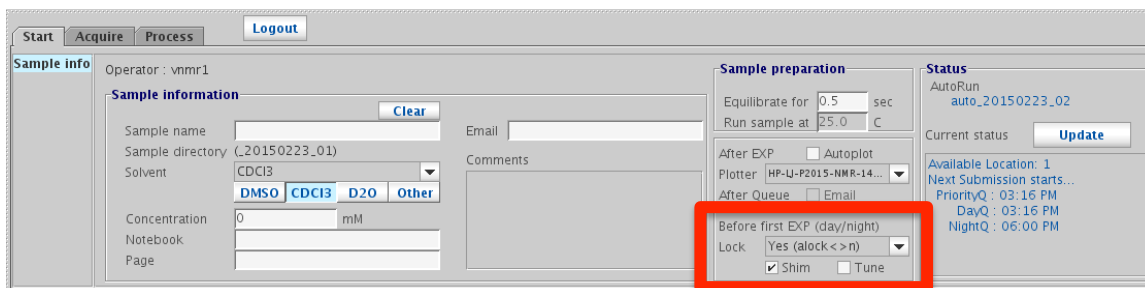


2. Click on **New study** in the Study Queue panel to go to the “Submit mode”. You will create ONE “New Study” for every sample you have.
3. In the Horizontal Panel, click **Start** tab to show the “Sample Info” page below.

Start → Sample Info page

- Fill out the **Sample name** box with a meaningful name for the sample (Letters and Numbers ONLY plus underscore _. No spaces and special characters).

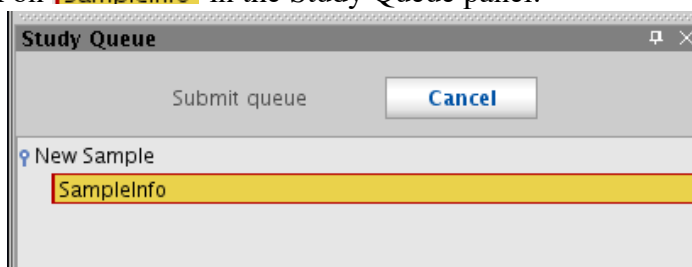
- Choose the Solvent  your sample is using.
- The rest of the text boxes are optional.
- **Tune the probe:**
 - When to check the  box:
 - When running any other nuclei than ^1H
 - When aqueous solvents are used
 - When salt concentration is high
 - When running variable temperature experiments



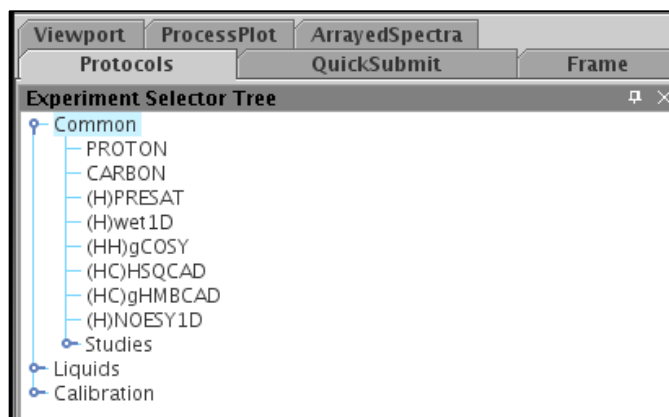

If you would like to do NMR on ^{13}C , ^{11}B , ^{19}F , ^{29}Si , ^{31}P , etc. please come talk to Shamon (CNSI Room 1528) for complete instruction.



If you want to change the information on the **Start** → **Sample Info** page later on, click on **SampleInfo** in the Study Queue panel:



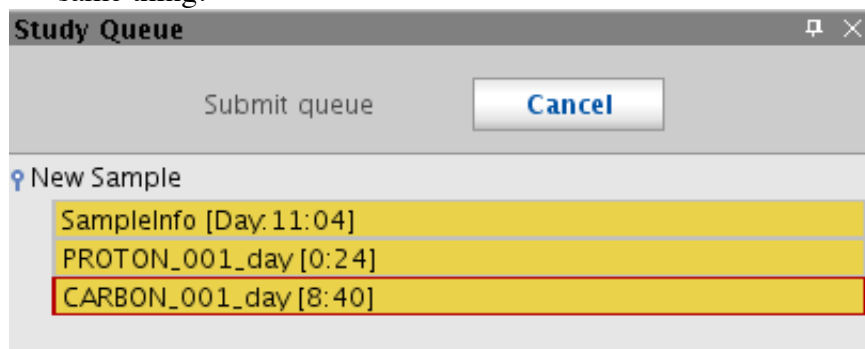
4. Select experiments with double-clicks from the lists under **Experiment Selector** in the vertical panel of VNMRJ window. To add NMR experiments to the **Study Queue**, double-click the desired experiments.



To delete an experiment from the Study Queue, right-click and select DELETE.



If other 1D NMR's and 2D NMR's are to be run, do basically the same thing.

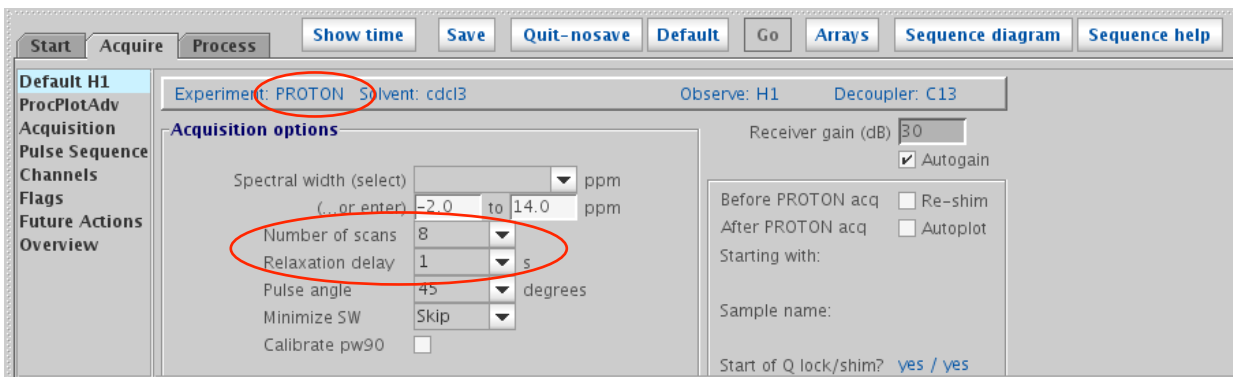


- Double click on node **PROTON_001_day [0:24]** in the **Study Queue** to load parameters for ^1H . If successful, you will see a new graph in the NMR Data

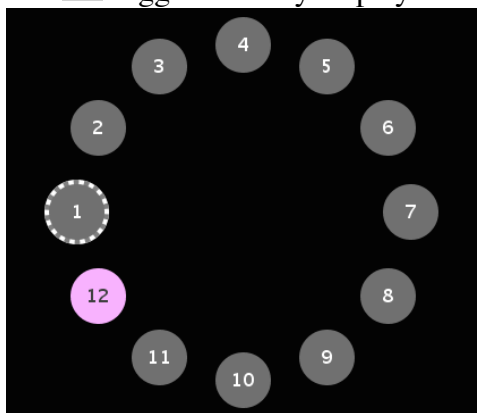
Display area and the pages associated with the **Acquire** tab in the Horizontal Panel, as shown below.

Change **Number of scans** and **Relaxation delay** as necessary by clicking on the **Acquire** tab and then on **Default H1**.

Acquire tab → Default H1 section



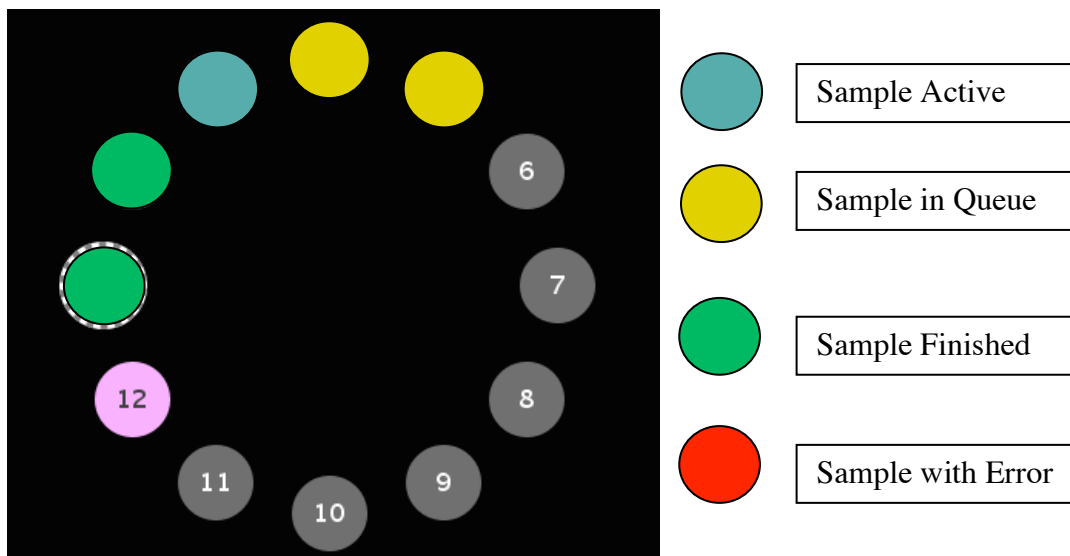
6. Do the same for ^{13}C : double click on node **CARBON_001_day [8:40]** in the **Study Queue** to load parameters for ^{13}C .
7. In the Horizontal Panel, click on the **Acquire** tab and then on **Default C13** to show the default ^{13}C parameters. Change **Number of scans** and **Relaxation delay** as necessary.
8. Click on  at the top left corner of the Graphics Display area and then on the numbered circle where the 1st sample is located (slot 1) in the tray display to have it selected: (:  and  toggles the tray display and NMR data display)



9. Click on  to start data acquisition. The auto-sampler will start changing samples by lifting up the CDCl_3 standard from the magnet, rotating counter-clockwise, and loading the sample in the 1st slot to the magnet. Followed by auto-tune, auto-lock, gradient-shim, and data acquisition.

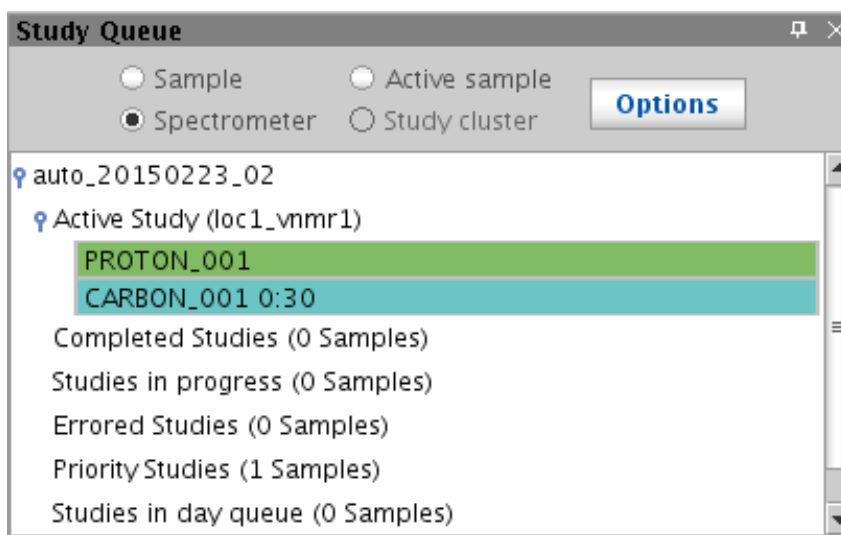


Color codes in the tray panel:



- To stop a running experiment and move on to the next one in the queue, use **Stop-Save-Resume** or **Stop-Discard-Resume** under the **Automation** → **Automation Controls** menu.
- To abort a whole automation run, use **Abort Automation** and then **Resume Automation** (or the machine will hang in the automation and nobody can use it) under the **Automation** → **Automation Controls** menu.

- While waiting for all experiments for Sample 1 to finish, you may setup the experiments for Samples 2, 3, etc. by clicking on  as necessary and then repeating steps 3 to 11 above to create a new Study Queue for a new sample.
- To view progress of experiments or process the finished experiments, look in the Study Queue window and make sure “Spectrometer” is checked:



 Same color codes as in the tray display:  finished,  running, and  waiting.

 You don't have to wait until the whole Study Queue (sample) finishes in order to process data. For example, you can process the Proton data while waiting for Carbon to end by double clicking the Proton node  PROTON_001.

 To toggle between the normal graphics display and the tray display, click on  or  near the top-left corner of the graphics display area.

14. At the end of the Automation Run, the sample tray will rotate multiple times to Sample Position #12. It will load the deuterated solvent sample into the magnet **automatically**.

VI. Finishing up

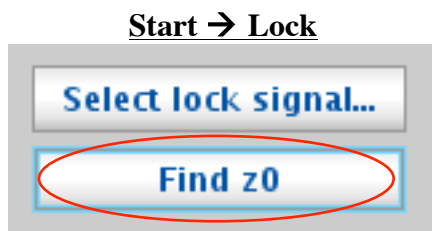


YOU ARE NOT FINISHED WITH THE SPECTROMETER UNTIL YOU DO THE FOLLOWING.

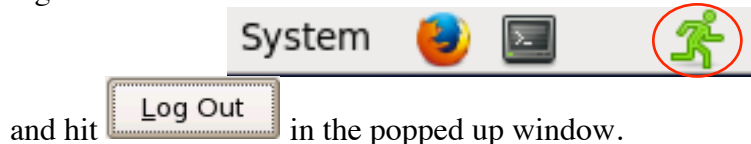
- a. Set solvent to CDCl_3 on the **Start** → **Standard** page as shown:

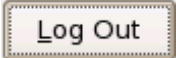


- b. Click on  on the **Start** → **Lock** page and wait for finish:



- c. Exit VNMRJ by clicking the X in the upper right of the screen.
- d. Log off your account: in the bottom left corner of screen click on the
logout button .



- and hit  in the popped up window.
- e. **Important:** On the logsheet, record your stop and duration times, and the spectrometer status. Report problems if any.
- f. Remove your samples from the lab and clean the space you have used.

VII. Data Processing Workstation

- Please go to NMR Processing Room (CNSI Room 1522) and use the computer titled “**Varian NMR Data Processing**”

1. Login
 - a. Type “**vnmrp**” as a username and input password “**4epp967**”
2. Click “**New Home_600MHz**” icon on the desktop to access files



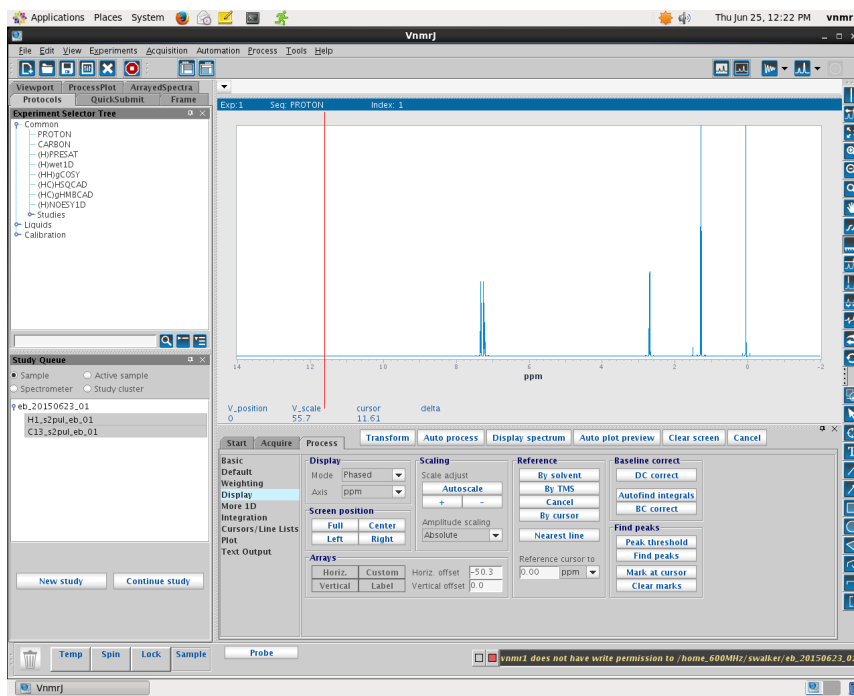
3. Open “**VNMRJ**” software on desktop

VIII. NMR Data Processing

- 1) Load your NMR file:



Choose “Open” → Your username → VNMRsys → Data → Your experiment → **Open the .FID file** ( Not the folder!)

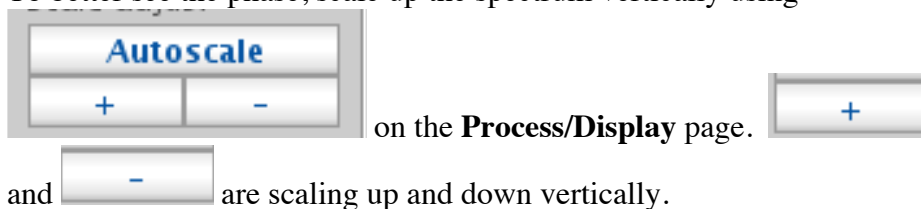


- 2) Click on  near the upper right corner of VNMRJ 4.2 to display the Spectrum and Spectrum Tool Bar. (Optional: you can click on  near the upper right corner to view the FID and the FID Tool Bar).

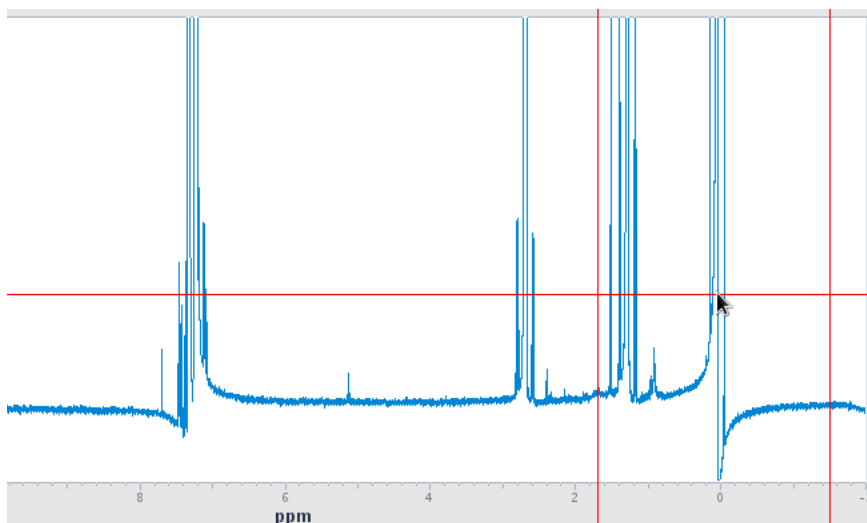
	Toggle 1 or 2 cursors
	Show Full Spectrum
	Reset to Full Display
	ZOOM IN
	ZOOM OUT
	ZOOM Mode
	Move Spectrum
	Integrate
	Show/Hide Axis
	Peel Threshold
	Find Peaks
	Display Integrals
	Phase Mode
	Redraw
	Return

- 3) Phase the spectrum:

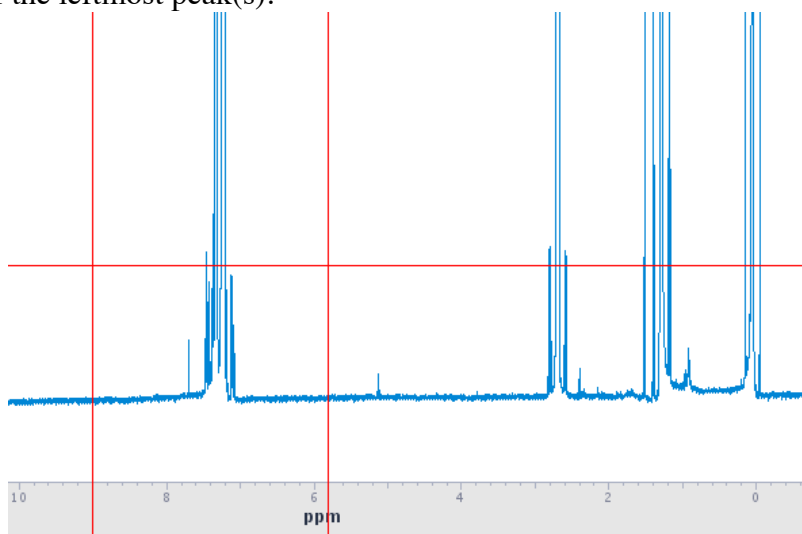
- a) To better see the phase, scale up the spectrum vertically using



- b) Click on  in Graphics Tool Bar and then on the rightmost region of peaks on the spectrum. The region (~2 ppm wide) will be marked with two vertical red lines and one horizontal red line as below:



- c) Place the mouse cursor on the horizontal red line between the two vertical lines, click-and-hold the right or left mouse button and move the mouse back and forth for fine or coarse phase adjustment, respectively, of the phase for the rightmost peak(s).
- d) Once the peak(s) are phased, click on the leftmost region of peaks (~2 ppm wide) and use the right or the left mouse button for phase adjustment of the leftmost peak(s).

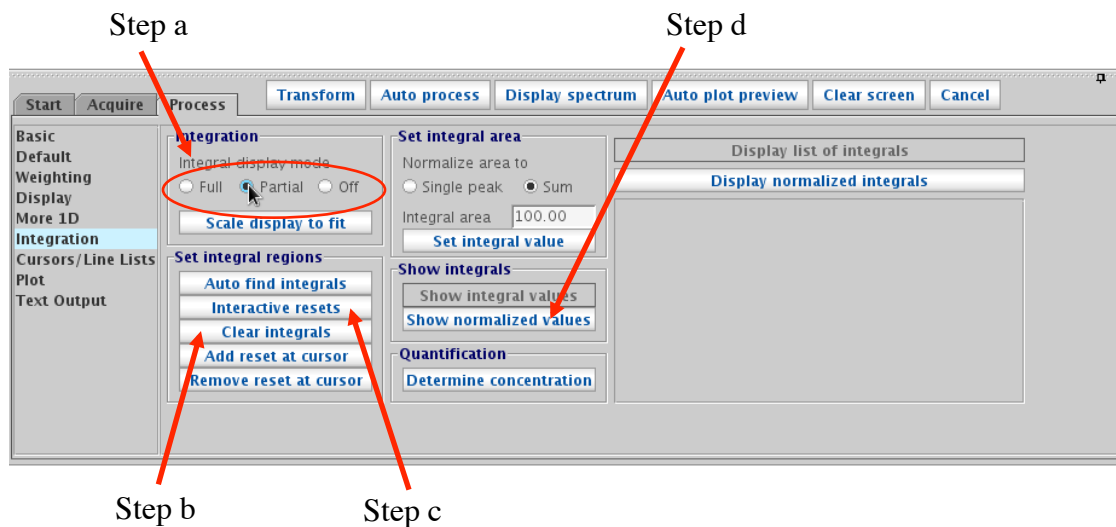


- e) Repeat steps b) to d) if necessary.

- f) After phasing, click on  in the Graphics Tool Bar to refresh spectrum display and on  on the **Process/Display** page to normalize the spectrum if necessary.

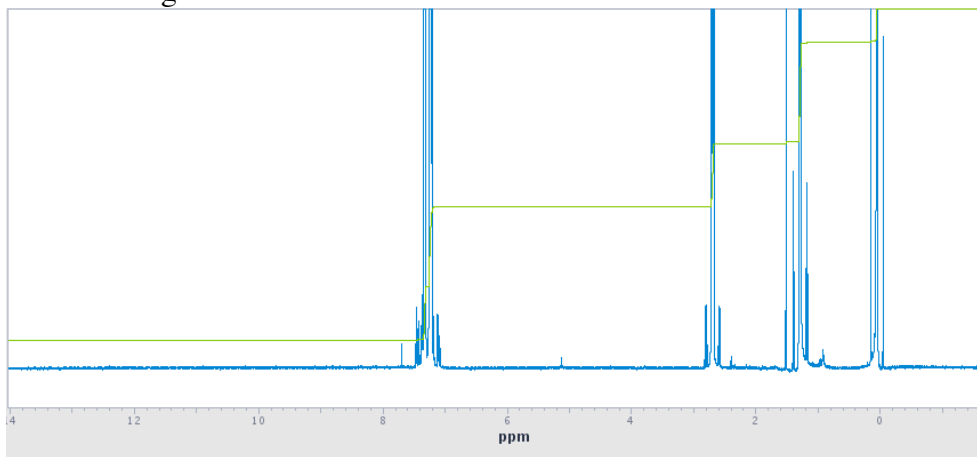
- 4) Baseline correction for accurate integration: on the **Process/Display** section, click on **DC correct** to correct the baseline DC offset and then on **BC correct** to get rid of the curvature or wobbling of the baseline.
- 5) Integration: click on **Integration** to go to the **Process/Integration** page:

Process/Integration page

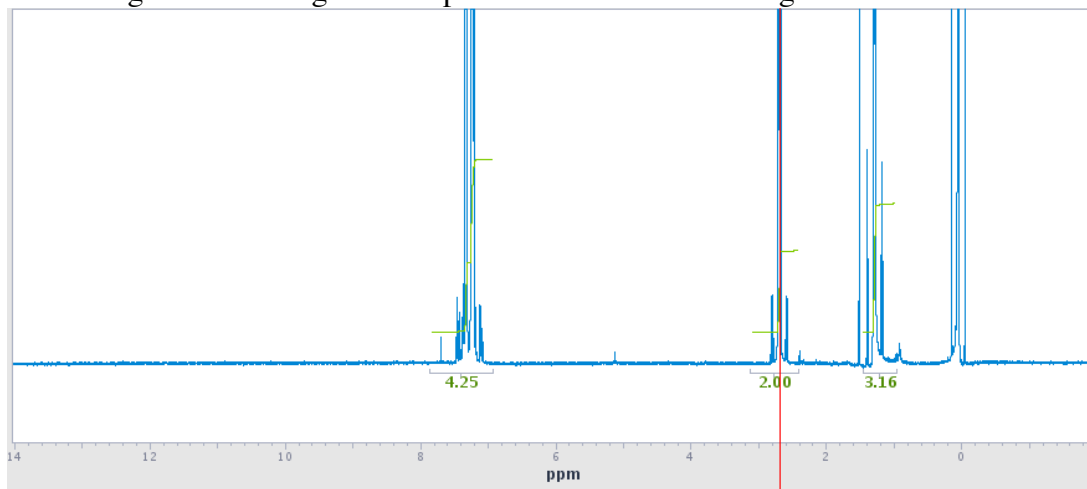


a. Make sure **Partial** is properly checked.

b. Click on **Clear integrals** to clear automatic integrals and all peaks are integrated as a whole:



- c. Click on **Interactive resets** and left-click on both sides of a region to be integrated. Repeat left-clicks on other regions of interest.



- d. To show integrals on spectrum, click on **Show normalized values**.



To calibrate the integrals to a single peak:

- Go to activate the single-cursor mode by clicking on in the Graphics Tool Bar so that you see a vertical red line on the spectrum. Place the cursor over the peak you want to calibrate.

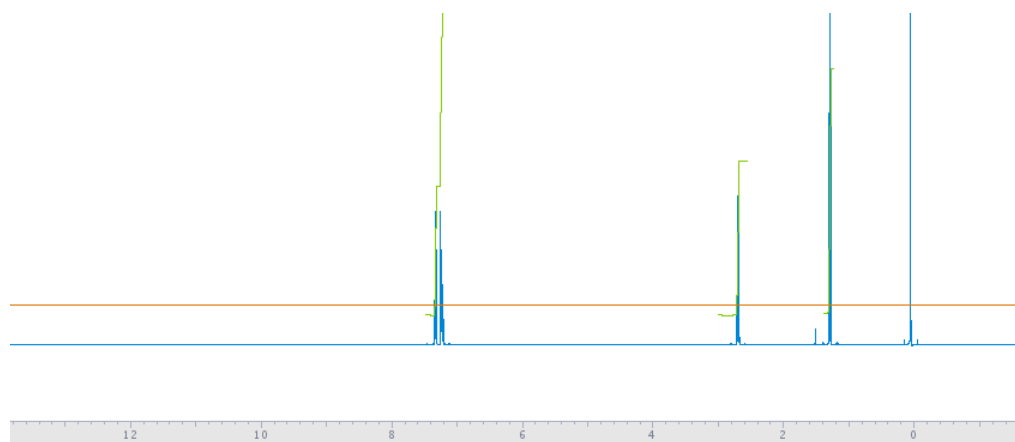
- #1 →
- #2 →
- #3 →
- #4 →

Set integral area
 Normalize area to
☒ Single peak ☐ Sum
 Integral area
Set integral value

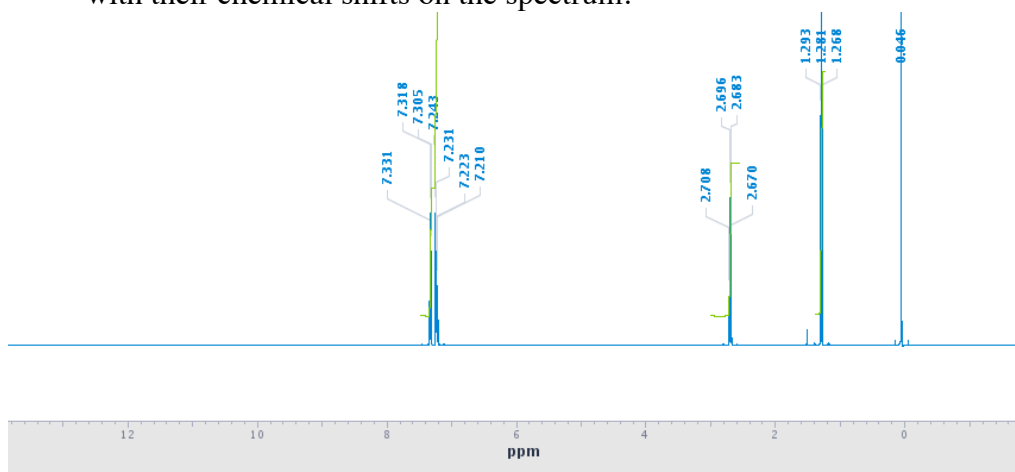
Show integrals
Show integral values
Show normalized values

6) Peak Picking:

- i. Click on **Peak threshold** on the **Process/Display** page to show the threshold (a horizontal yellow line) on the spectrum. Only those peaks taller than the threshold will be picked up.



- ii. Place the mouse cursor on the yellow line and use the left-mouse-button to drag the line up or down.
- iii. Click on Find peaks on the **Process/Display** page to label peaks with their chemical shifts on the spectrum.



IX. Appendices

1. Requirements for Access to the MRL NMR at CNSI

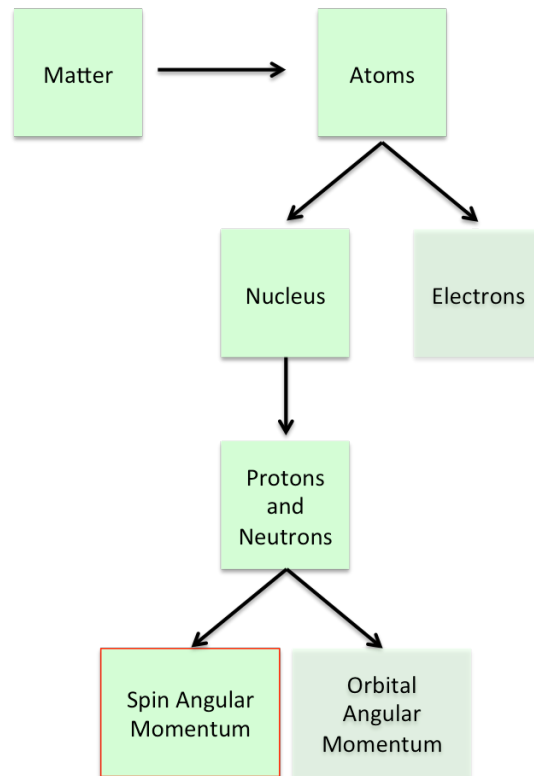
You have to pass the mini quiz within one month after training in order to be qualified for access to the NMR facility of MRL, which includes:

- Key Card for Lab & Building:
 1. Pass the MRL safety training;
 2. Fill out the CNSI access form:
http://www.cnsi.ucsb.edu/facilities/building_services/access/access_application.pdf
 3. Take the form to Sylvia in 2066G, MRL
- Web Scheduling Account
- NMR Account

These requirements apply to both on- and off-campus users.

X. NMR Basic Principles

1. Spin



*Spin is a quantum mechanical phenomena that has no physical analog in classical physics. However, it will be helpful to visualize it as a small bar magnet that precesses about an axis.

*The existence of spin angular momentum is inferred by experiments, such as the Stern-Gerlach experiment, in which particles are observed to have angular momentum that cannot be solely accounted for by orbital angular momentum alone.

*Electrons, protons, and neutrons all have a value of spin $\pm \frac{1}{2}$.

2. Common NMR Nuclei

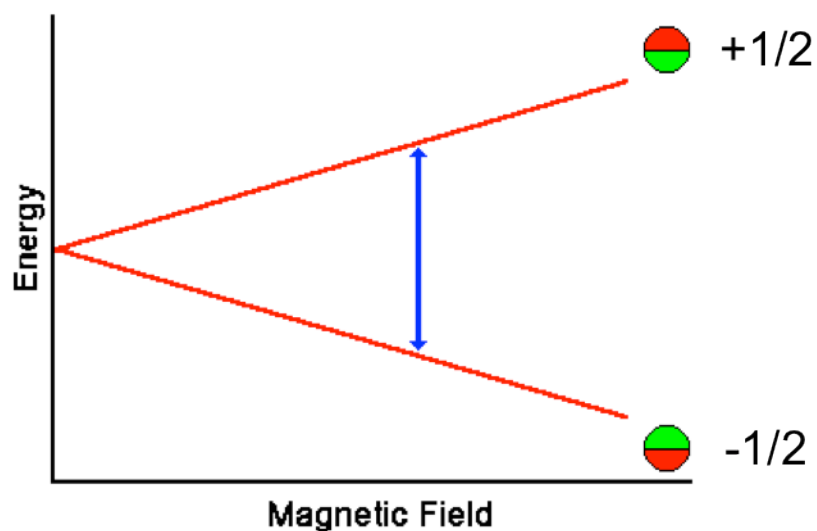
Nuclei	Unpaired Protons	Unpaired Neutrons	Net Spin	γ (MHz/T)
^1H	1	0	1/2	42.58
^2H	1	1	1	6.54
^{31}P	1	0	1/2	17.25
^{23}Na	1	2	3/2	11.27
^{14}N	1	1	1	3.08
^{13}C	0	1	1/2	10.71
^{19}F	1	0	1/2	40.08

Larmor Frequency Equation:

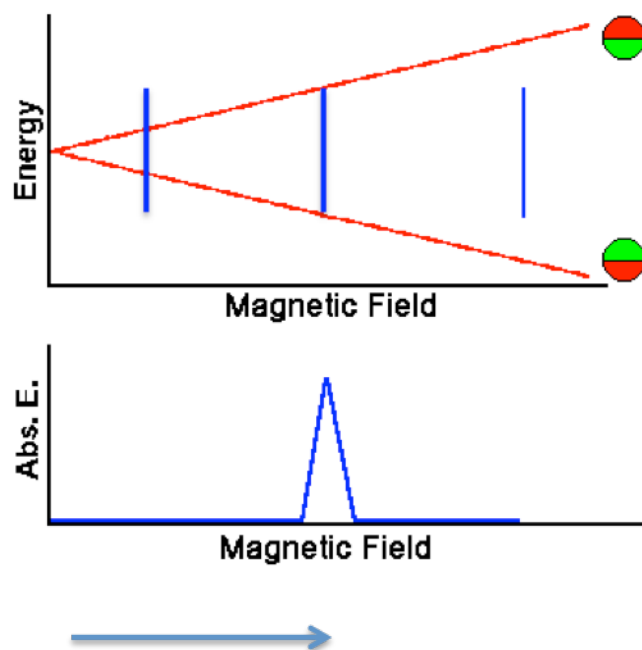
$$\nu = \gamma B_o$$

where γ is the gyromagnetic ratio (specific to each nuclei) and B_o is the magnetic field strength

3. Energy Level Diagram

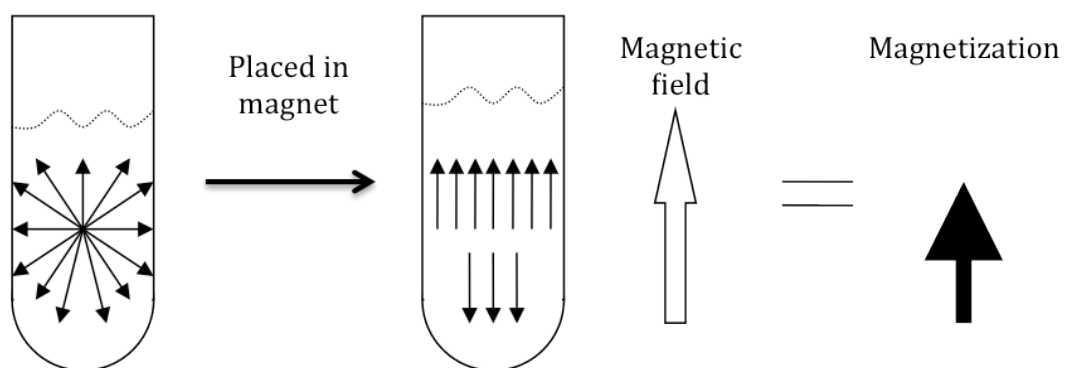


4. cw NMR



5. Magnetization

Alignment of nuclei in a magnetic field



6. Pulsed NMR, Relaxation, and Detection

